

Flow Cytometry & Immunofluorescence (Physics and Chemistry)

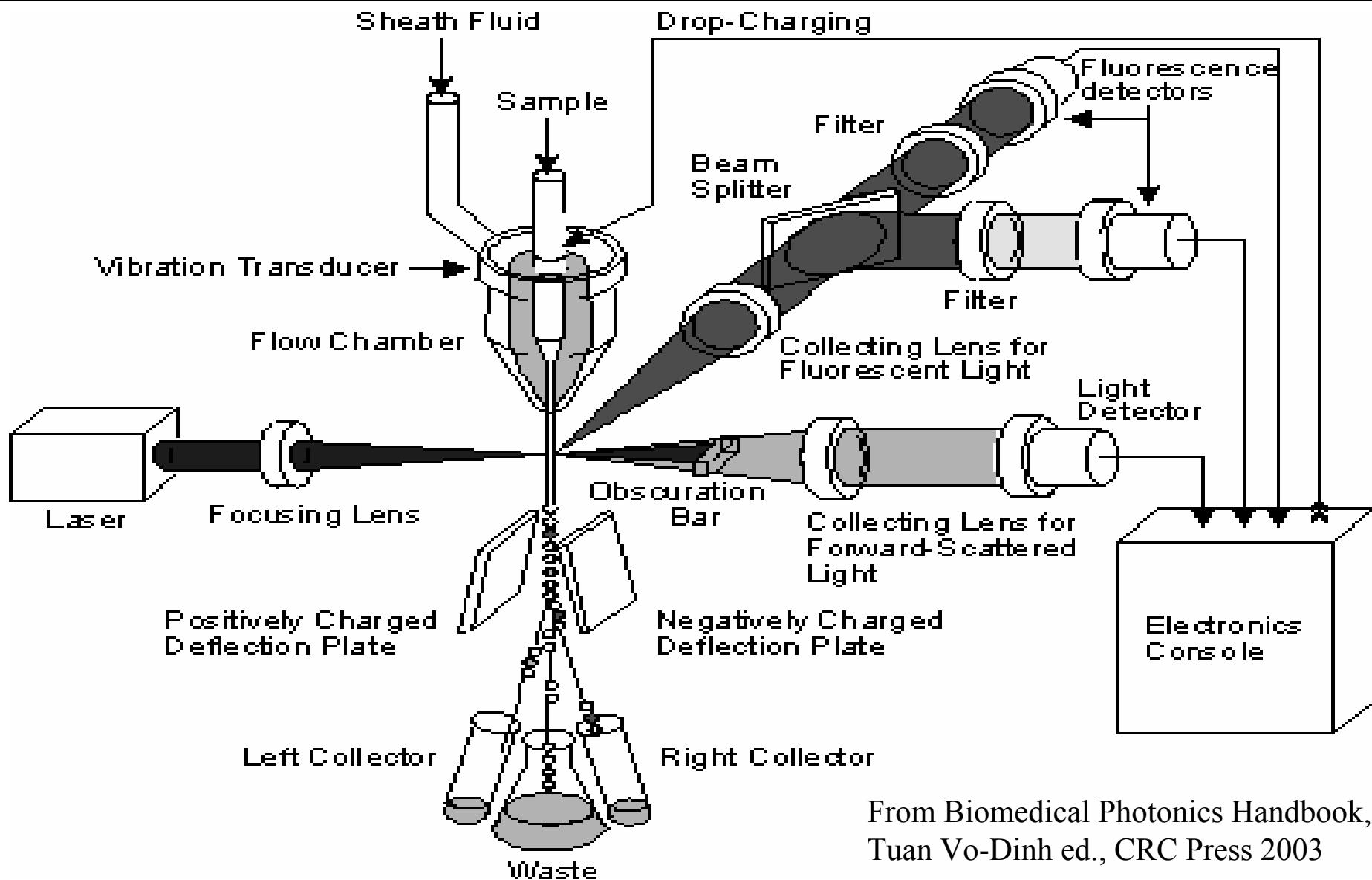
Presented by:
Diether Recktenwald, PhD

Contact email:
diether_recktenwald@bd.com or diether@att.net

BD Biosciences
Clontech
Discovery Labware
Immunocytometry Systems
Pharmingen

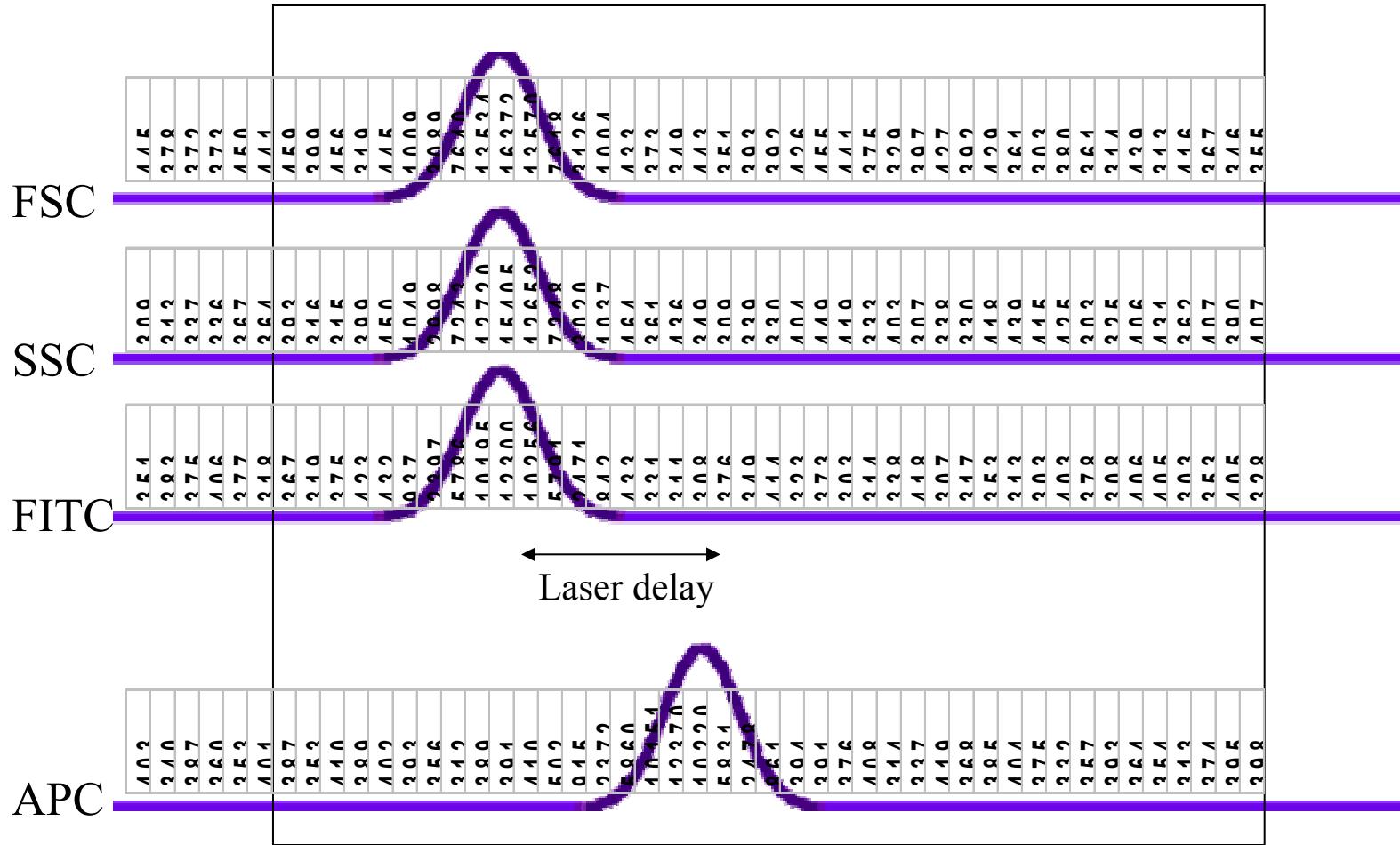


Flow Cytometer



From Biomedical Photonics Handbook,
Tuan Vo-Dinh ed., CRC Press 2003

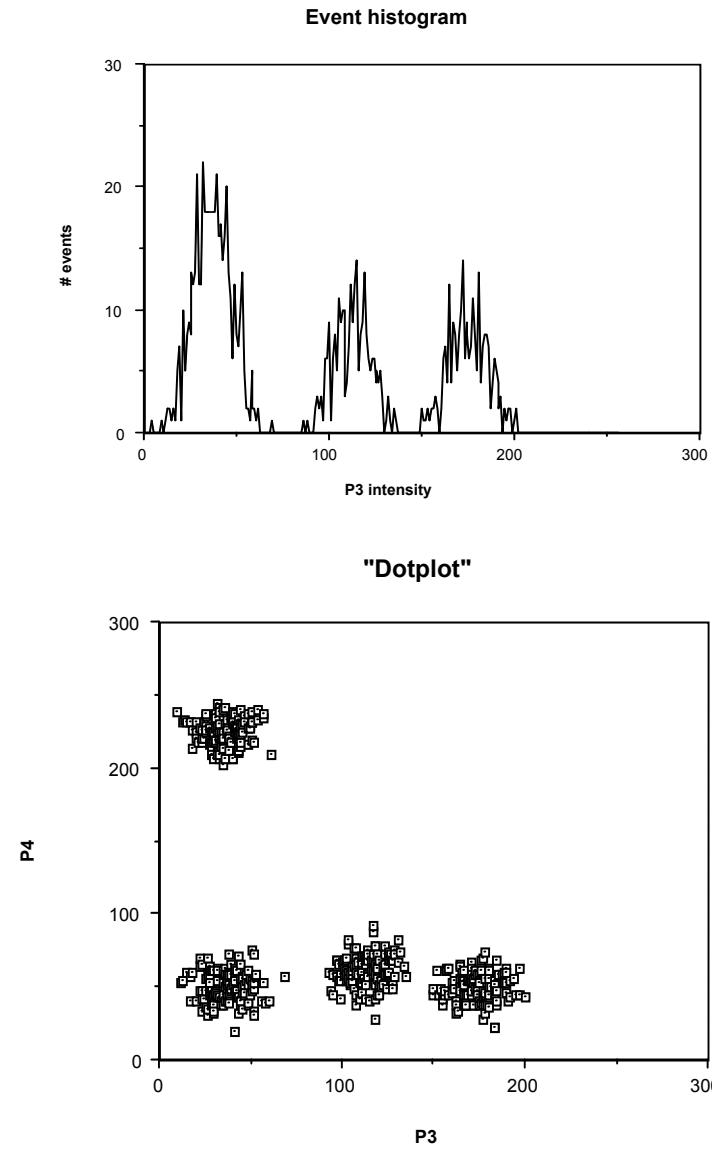
Numbers in Memory



Flow Cytometry Data Analysis

Cell	P1	P2	P3	P4	P5	Pop#
1	242	135	704	175	612	1
2	146	132	690	178	566	1
3	269	147	89	206	580	3
4	442	143	399	250	255	4
5	212	167	155	926	526	2
6	269	2	659	207	575	1
7	204	232	112	171	679	3
8	152	74	160	828	532	2
			...			
9997	215	119	138	936	662	2
9998	244	50	72	261	543	3
9999	214	137	174	1014	597	2
10000	312	87	110	904	560	2

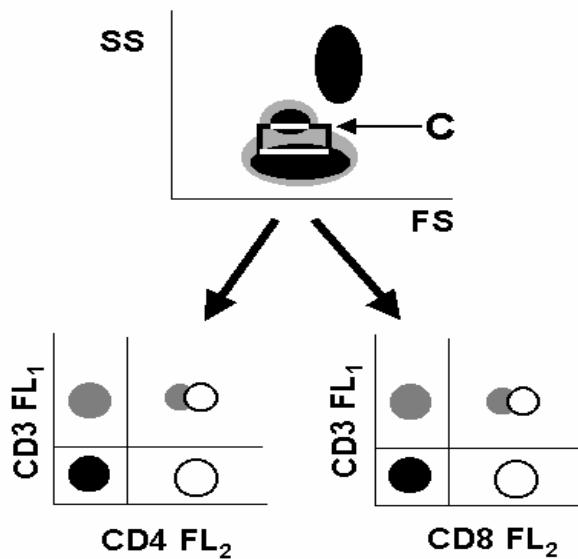
Note: more than 12 parameters
in advanced FACS systems



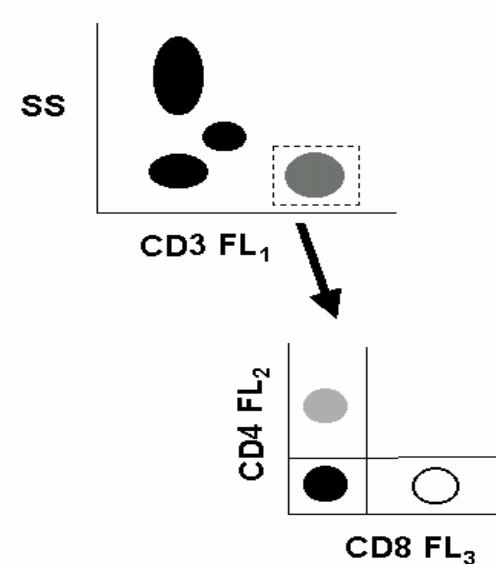
“Gating”

GATING STRATEGIES BASED ON INTRINSIC AND EXTRINSIC CELL ATTRIBUTES

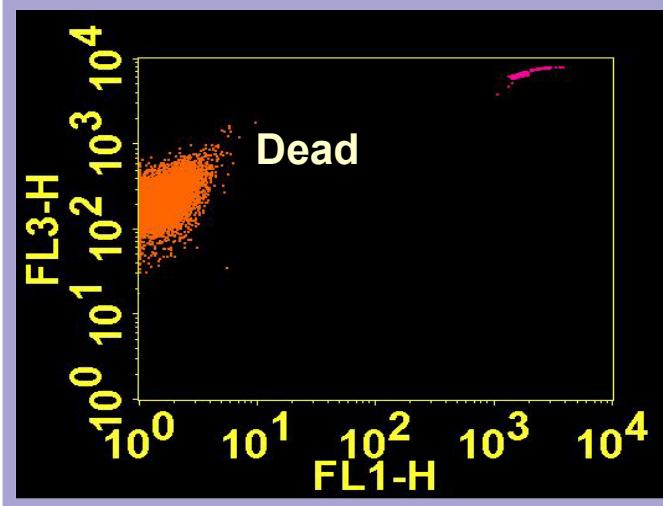
A



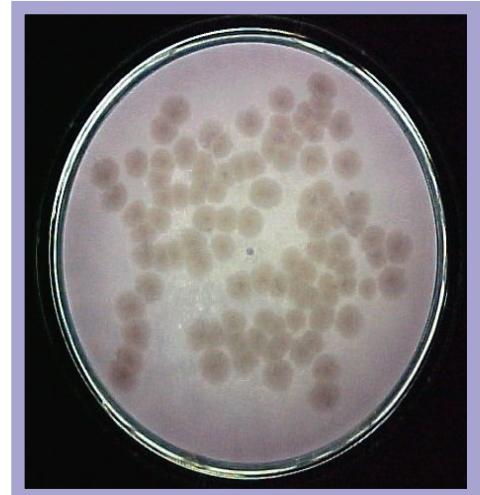
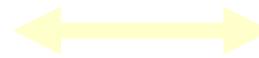
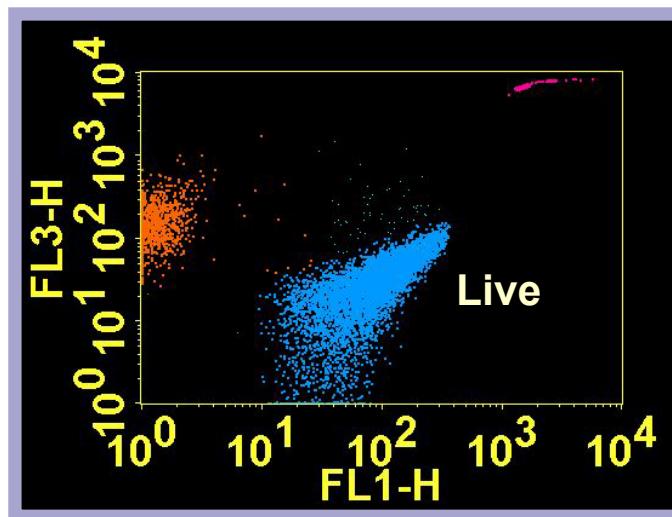
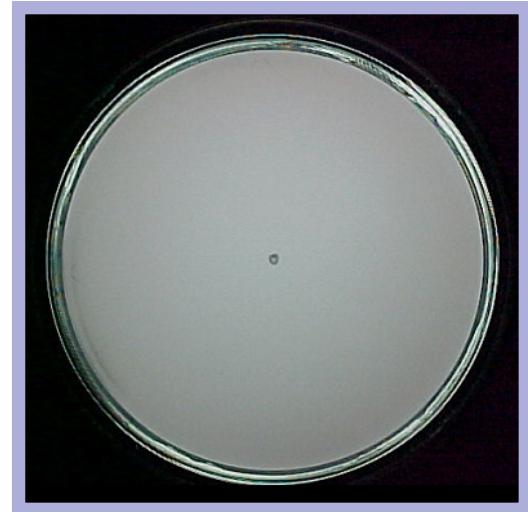
B



Cell Sorting for Functional Studies



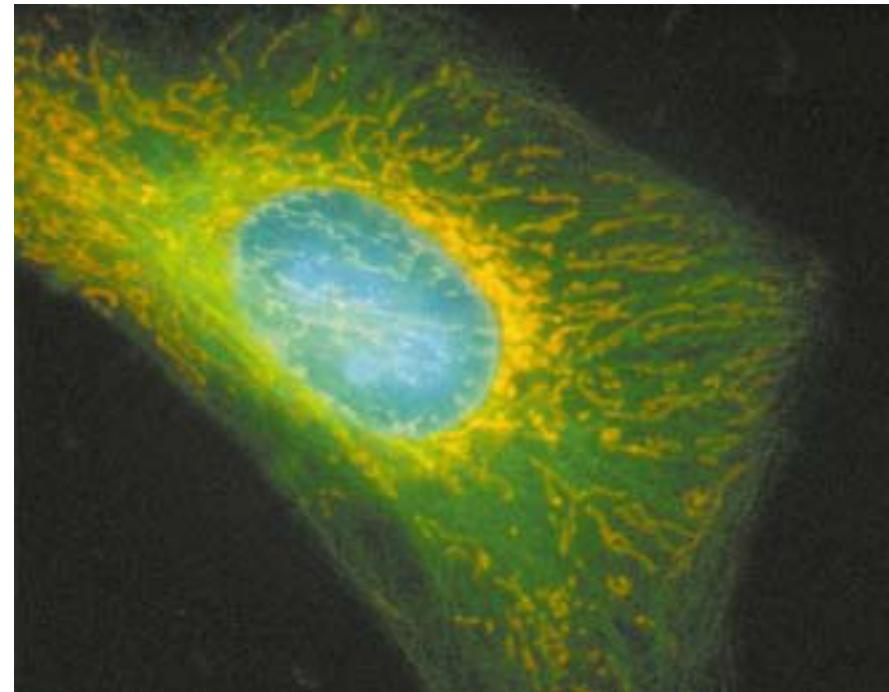
**Flow
Sorting
and
Plate
Detection**



Parameters For Cell Analysis by Flow Cytometry

Analyse and Sort based on:

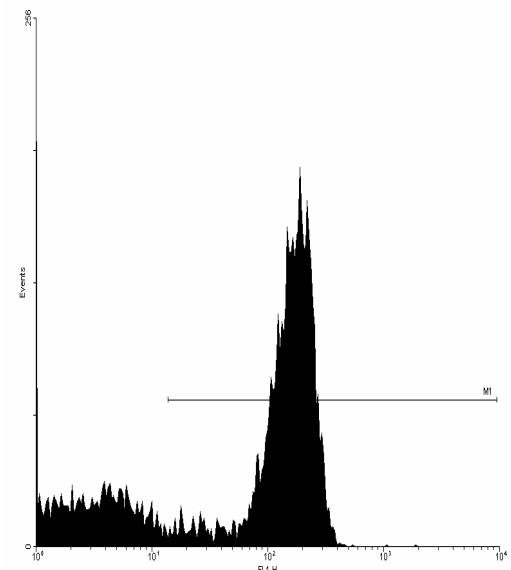
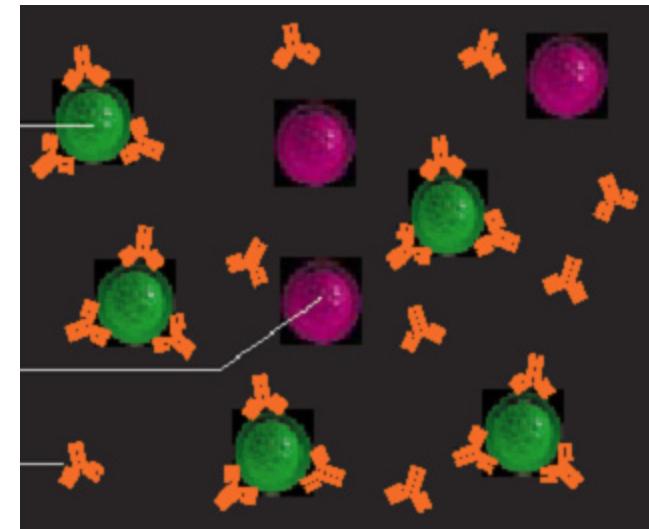
- light scatter
- immunofluorescence
- fluorescent in-situ hybridization
- DNA content
- transfection with fluorescent proteins
- protein content
- auto-fluorescence
- enzyme activity
- pH
- redox potential
- other components detectable by fluorescence



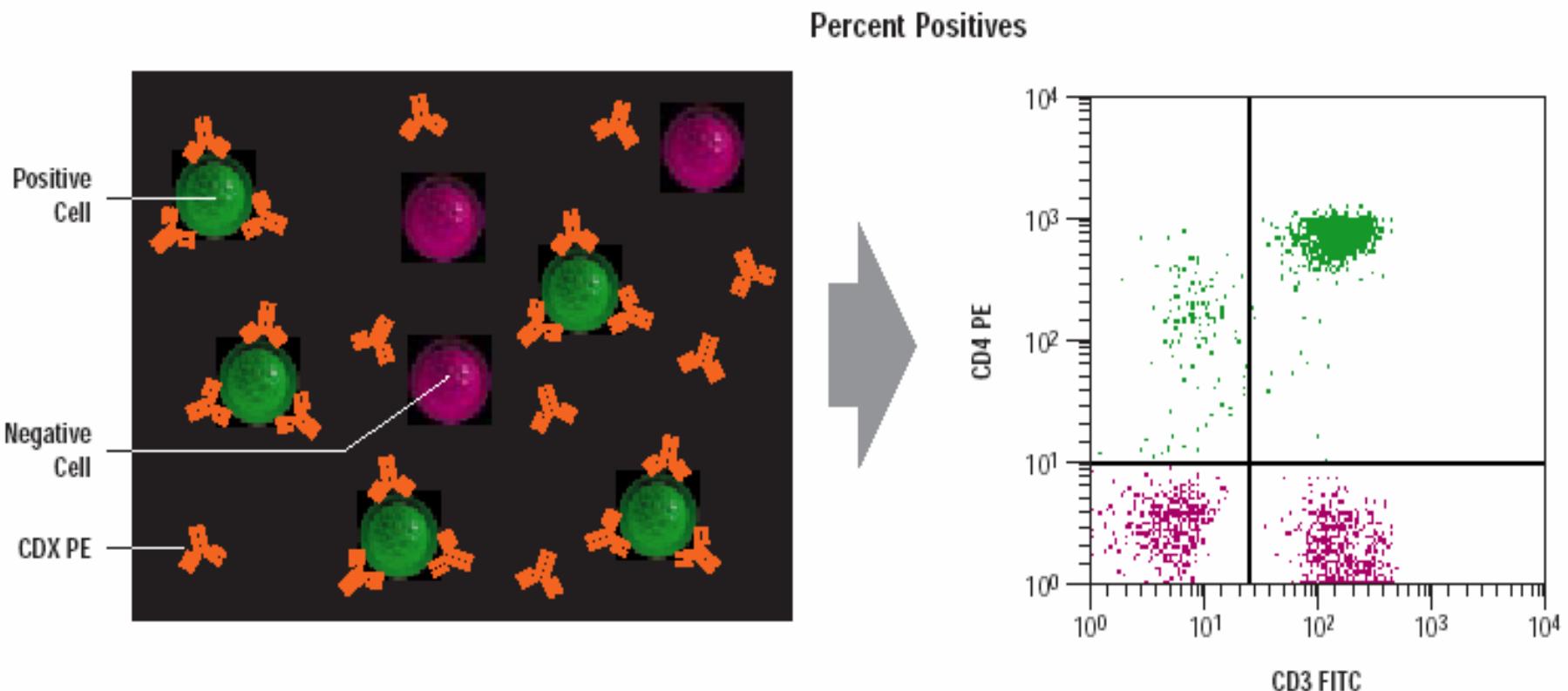
HeLa cells transfected with fluorescent protein vectors for nuclei, mitochondria and tubulin.

Immunofluorescence

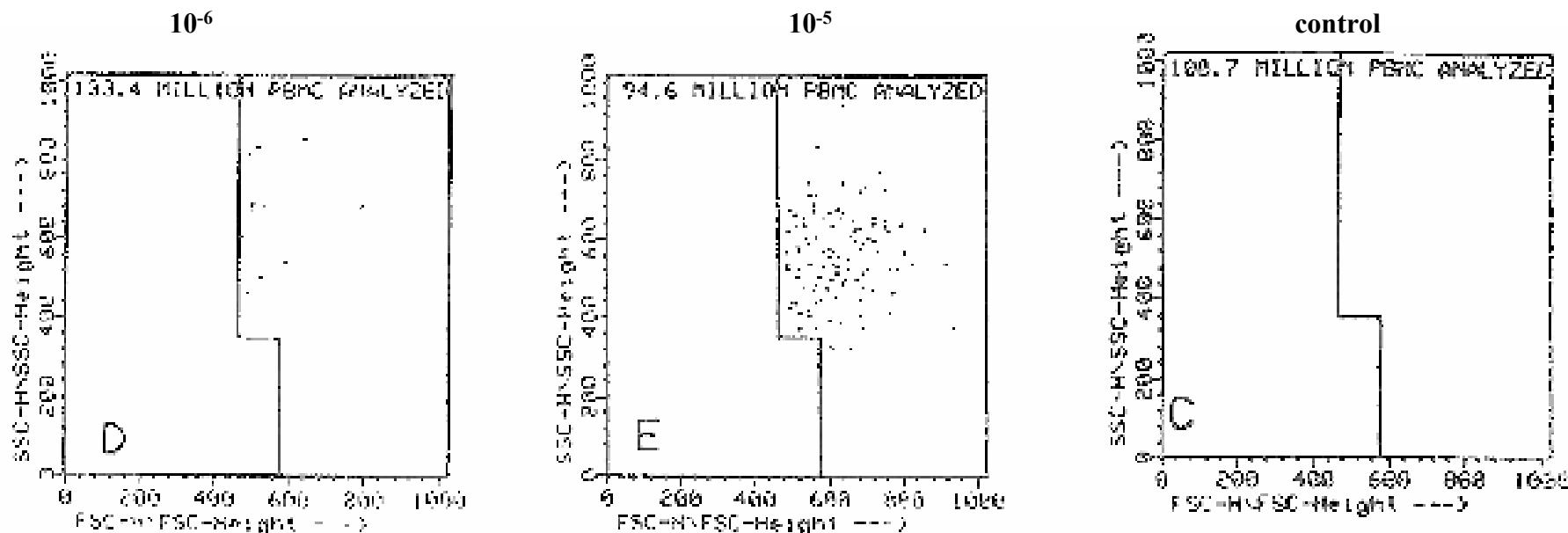
- Sample conditioning
 - Disaggregation of tissues
 - Pre-enrichment
- Reaction of sample with reagent
 - Direct or indirect immunofluorescence
 - Wash or no-wash
- Multi-color fluorescence measurement
- Data analysis



Immunofluorescence Data (1)



Immunofluorescence (1) Limit of Detection for Rare Cells



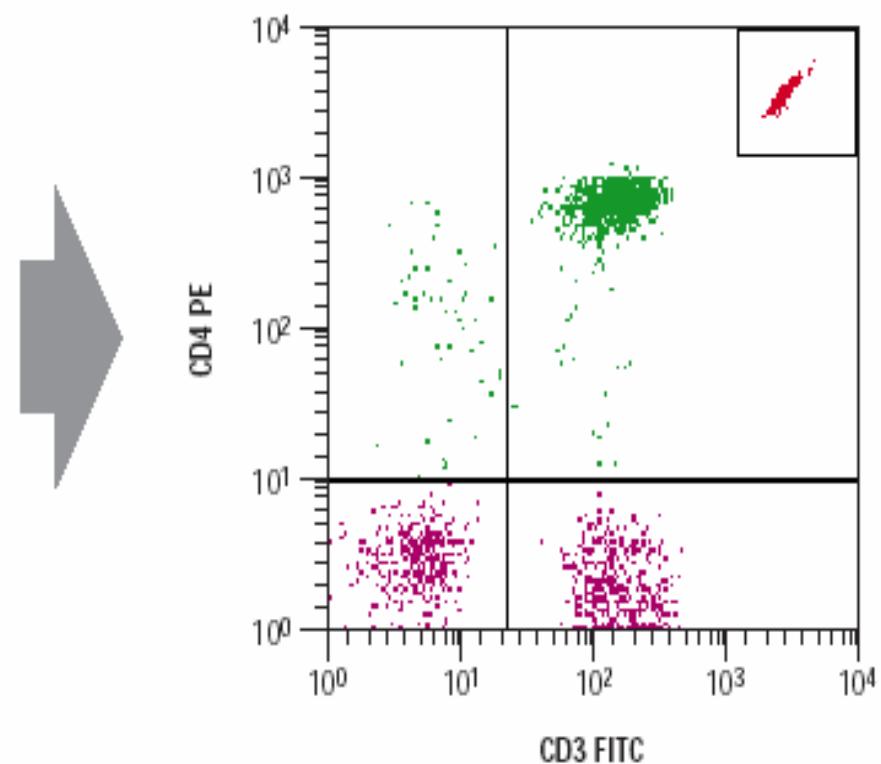
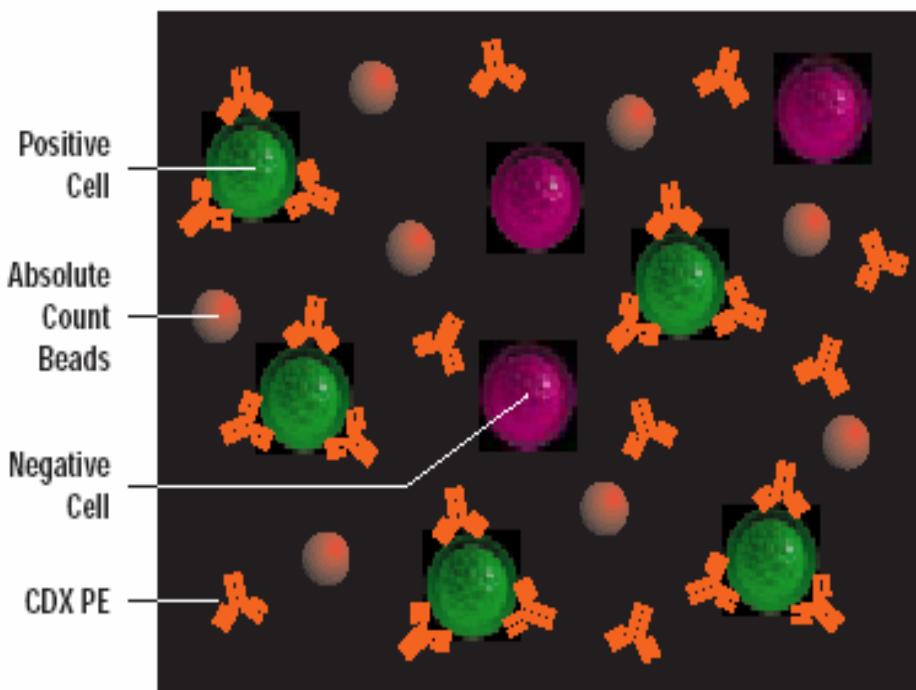
Routine $>0.2\%$

Optimized instrument $>0.01\%$

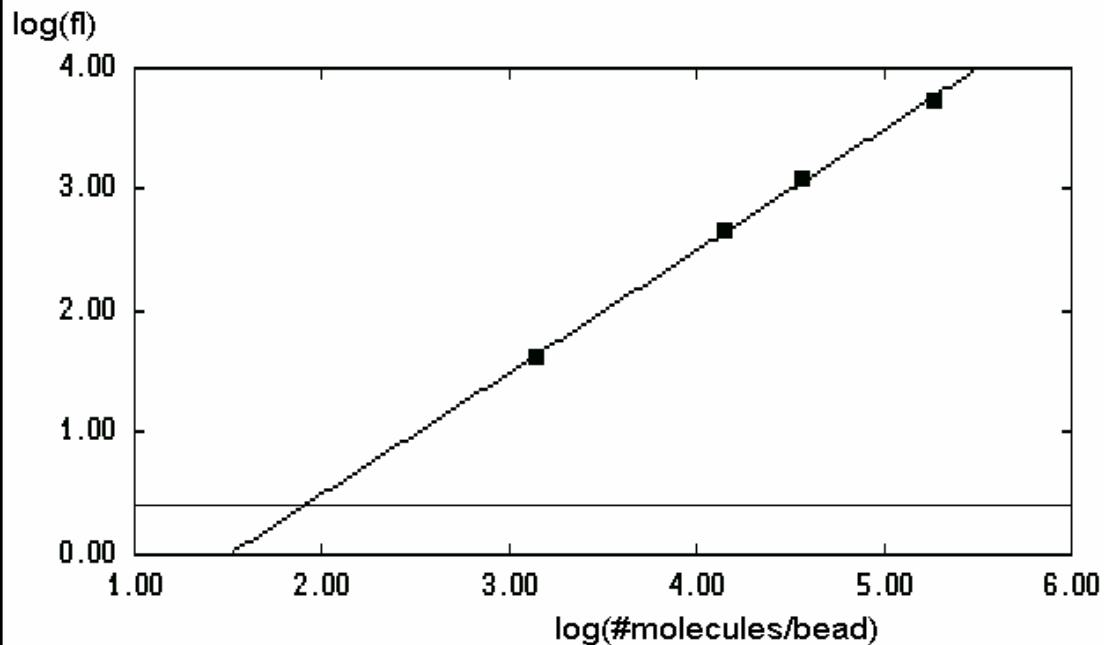
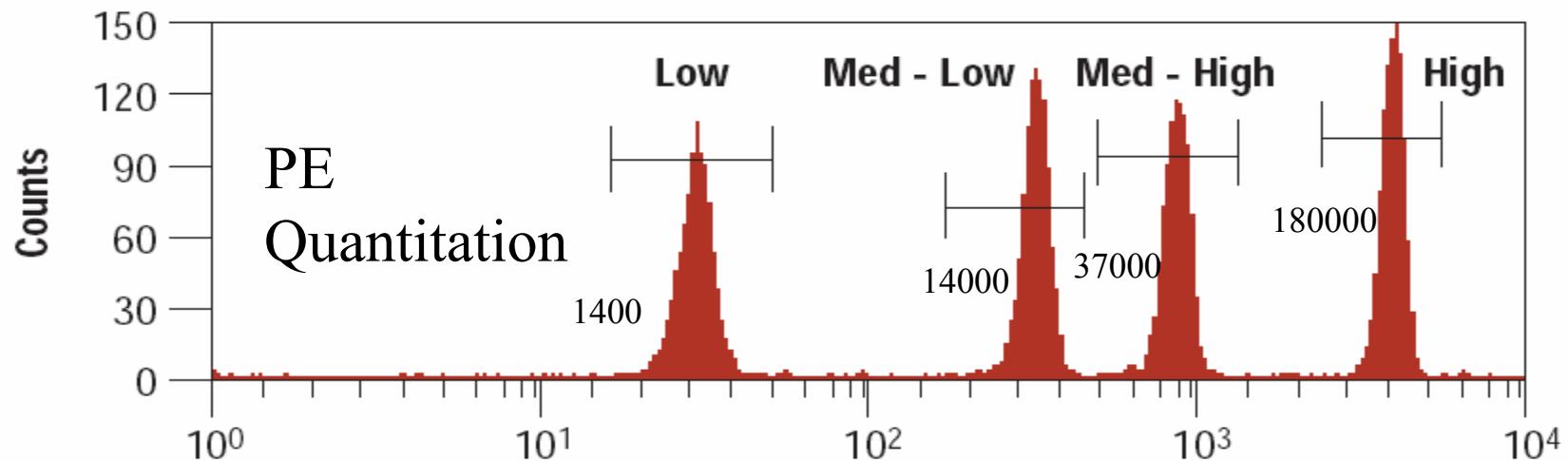
Optimized system $>10^{-7}$

Immunofluorescence Data (2)

Absolute Counts — Cells per μL



Immunofluorescence Data (3)

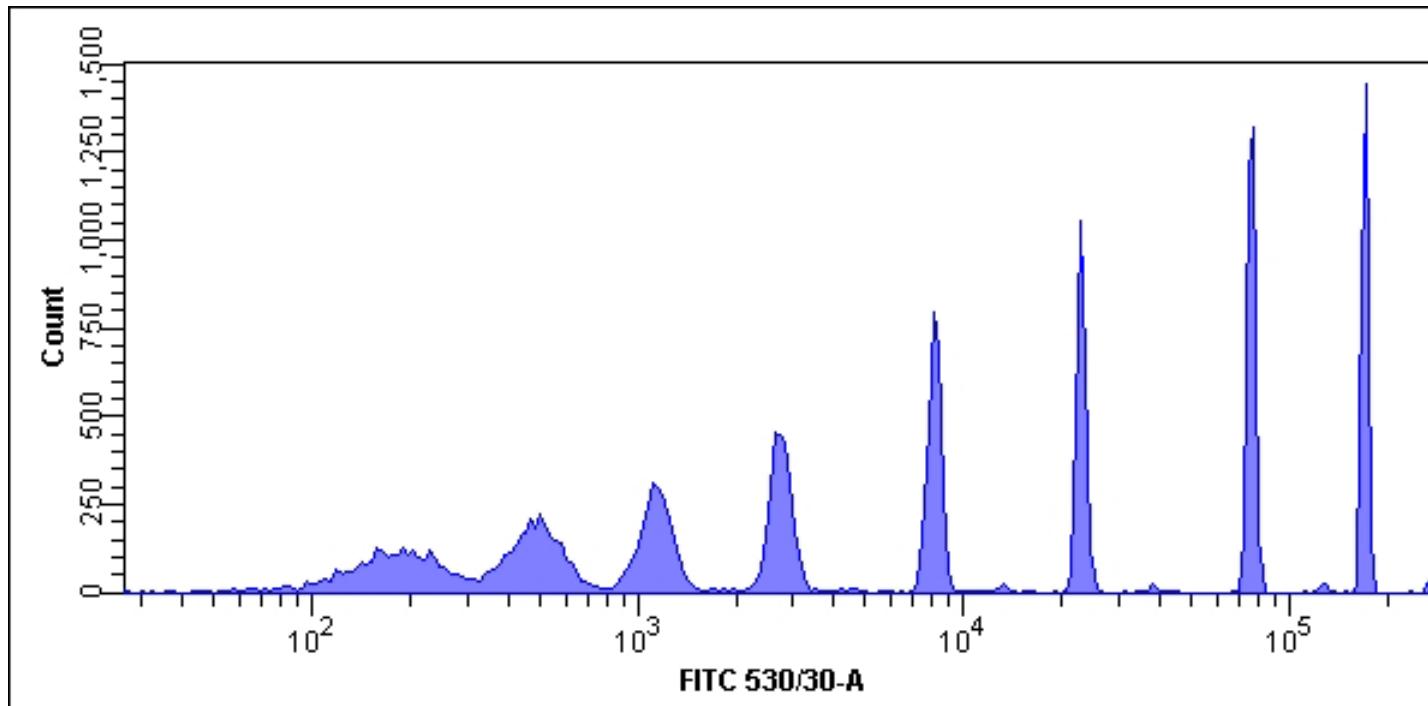


Molecule	#/cell
CD3	8.1×10^4
CD4	5.9×10^4
CD8	1.4×10^5
CD11a	2.7×10^4
CD16	7.9×10^4
CD18	3.1×10^4
CD45	1.9×10^5

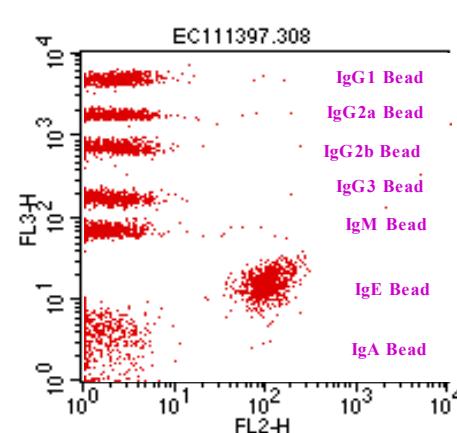
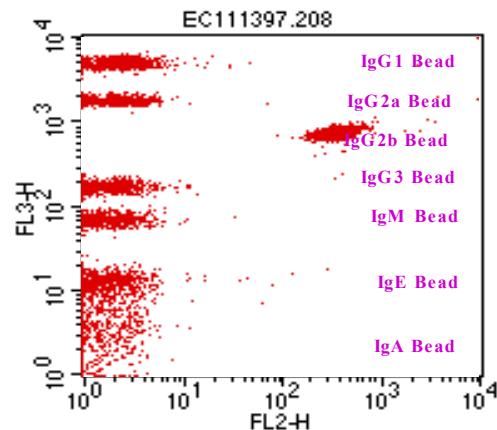
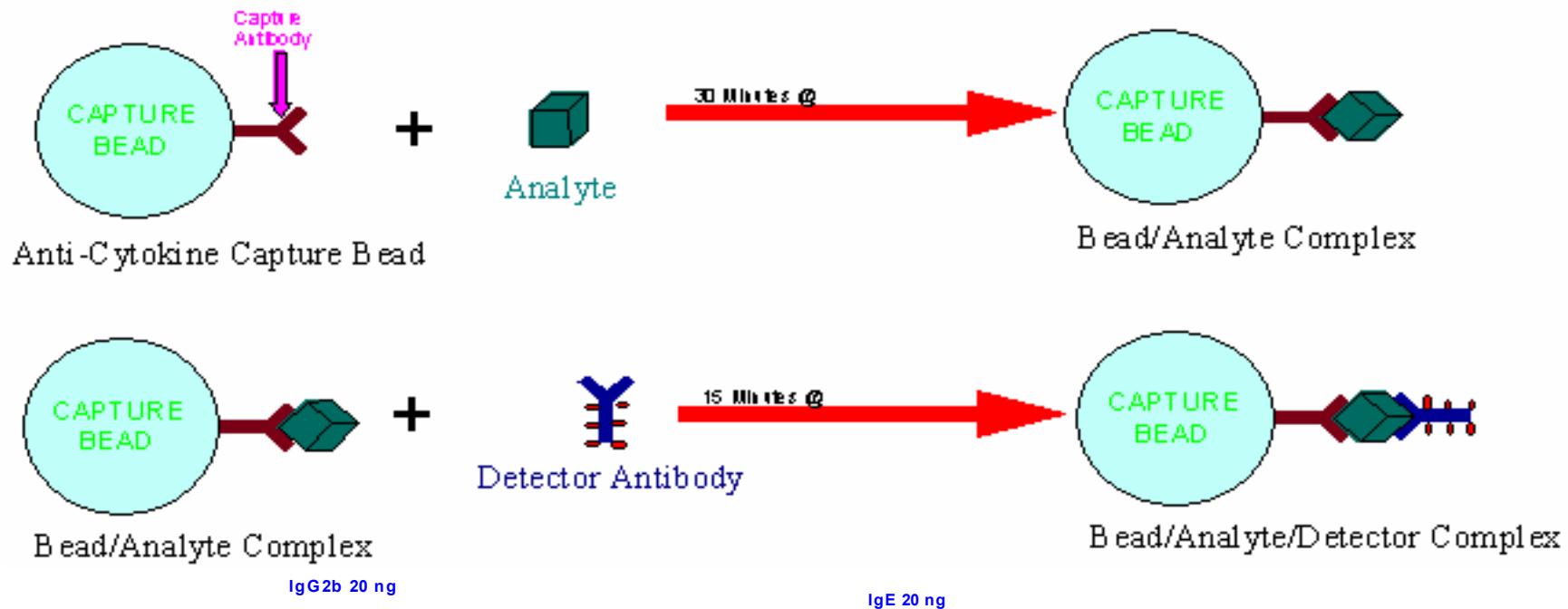
From:

Appendix A, Cell Separation Methods
and Applications. Marcel Dekker 1998.
Recktenwald D and Radbruch A, eds.

Sensitivity



Microsphere-based assays for soluble analytes



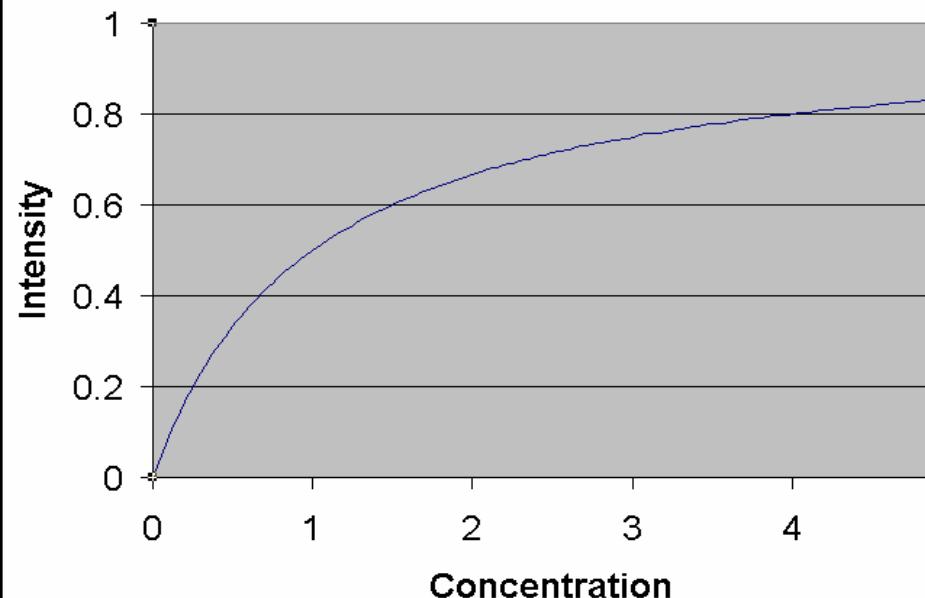
Dr. Rudi Varro, BD Biosciences

Immunofluorescence Issues

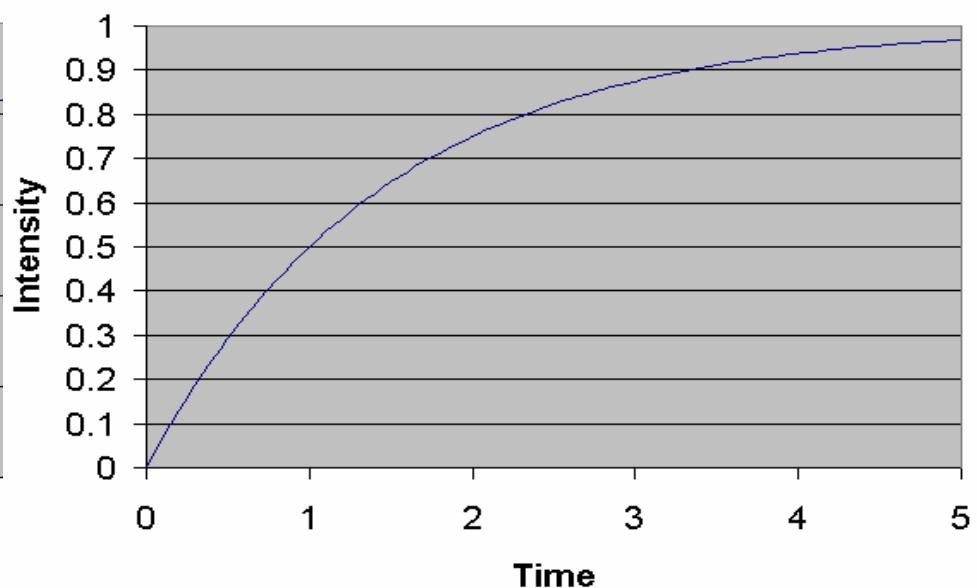
- Label selection (sensitivity and compensation)
- Photobleaching (especially energy transfer conjugates)
- Environment sensitive fluorescence (i.e.FITC)
- Fixation
- Dead cells (PI, EMA)
- Reagent equilibrium binding
- Binding kinetics
- “Non-specific” reactions

Ligand binding (1)

Equilibrium Binding

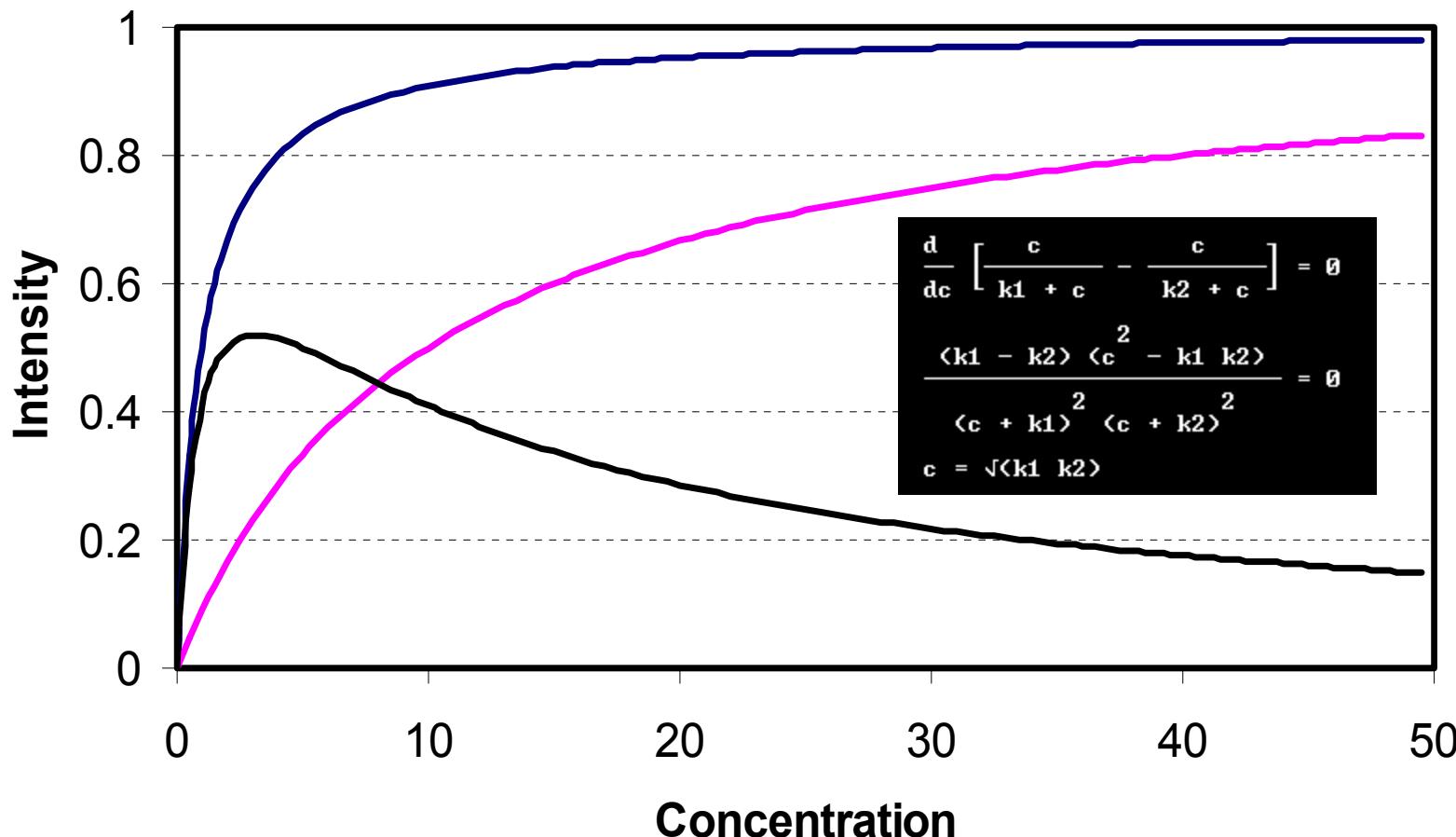


Kinetics



Ligand binding (2)

Effect of "non-specific" binding

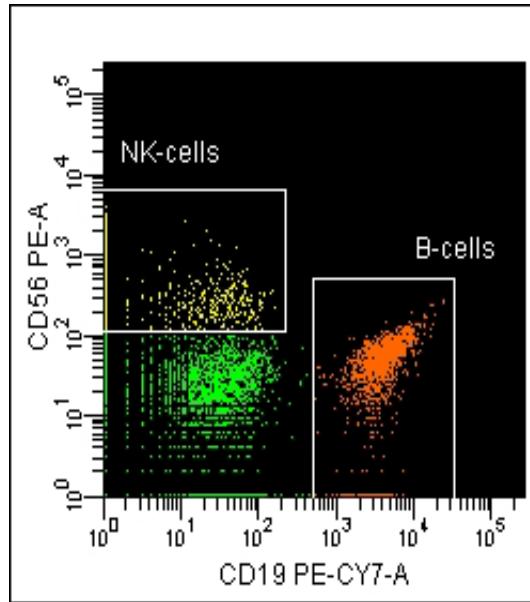
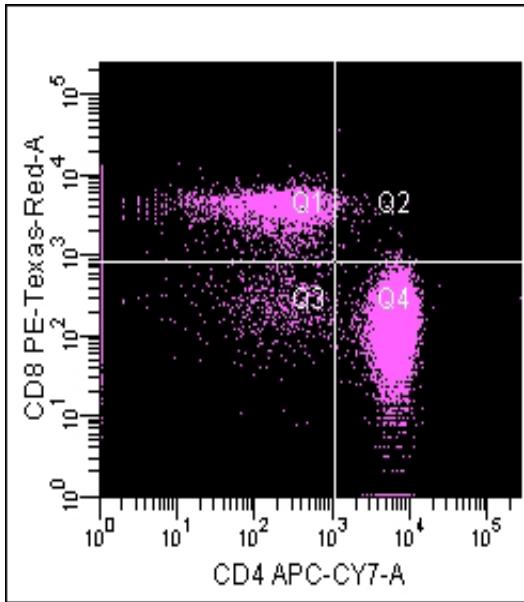
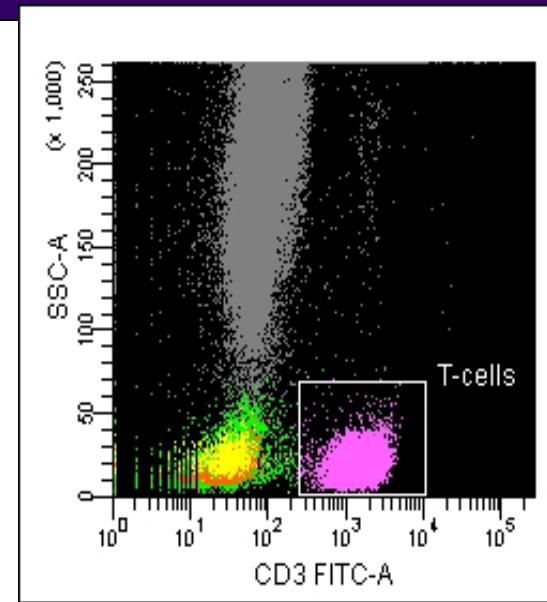
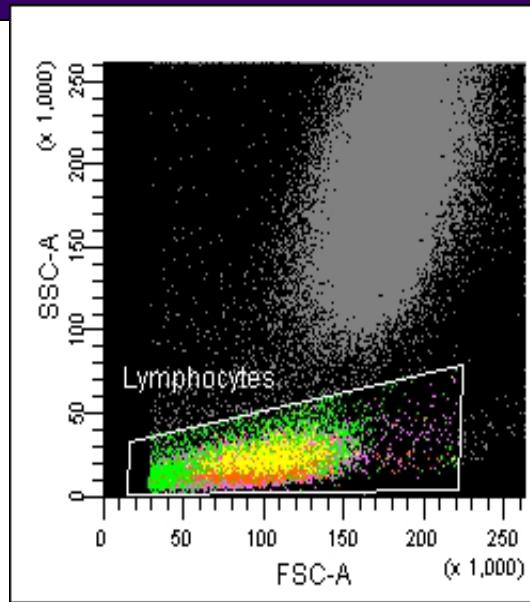
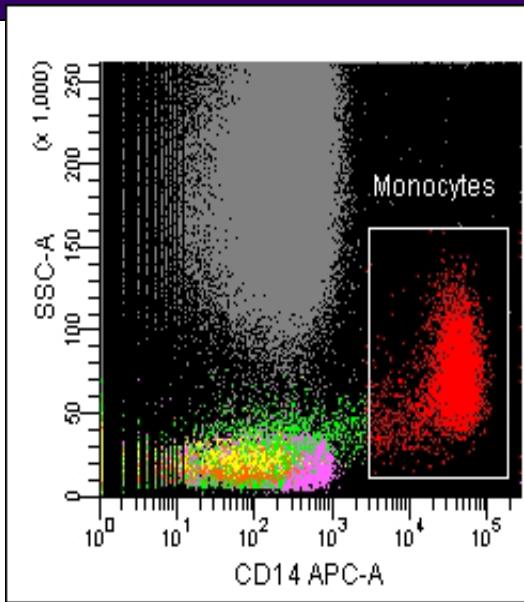


Immunofluorescence Multi-color Analysis

DYE	Excitation with		Emission (nm)	Extinction Coefficient (cm ⁻¹ M ⁻¹)	Quantum yield	MW (Da)
	488	635				
	1 st laser	2 nd laser				
FITC	X		519	67,000	0.71	389
PE	X		578	1,960,000	0.68	240,000
PerCP	X		675	na	1	35,000
ECD	X		613	na	na	250,000
PE-Cy5	X		675	1,960,000	na	241,000
APC		X	660	700,000	0.68	104,000
Cy5		X	670	250,000	0.28	792
APC-Cy7		X	767	700,000	na	105,000

From Biomedical Photonics Handbook, Tuan Vo-Dinh ed., CRC Press 2003

Six color example



- CD3 FITC
- CD56 PE
- CD8 PE-Texas Red
- CD19 PE-Cy7
- CD14 APC
- CD4 APC-Cy7

End

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